



¹H and ¹³C NMR conformational properties calculated by Spartan 18, and 400/125.77 MHz experimental data of methyl β-orcinolcarboxylate and atranorin, a comparative study

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ABSTRACT

¹H and ¹³C NMR properties calculated by Spartan 18 and 400/125.77 MHz experimental data of methyl β-orcinolcarboxylate and atranorin, a comparative study

The structural parameters of methyl β-orcinolcarboxylate and atranorin were determined by B3LYP with the base set 6-31G*. The results of the optimized molecular structure are presented and compared with the available X-ray data of the molecules. A comparative analysis of the experimental ¹H and ¹³C NMR spectra and calculated values is exposed and discussed. The theoretical quantum calculations resulted a good predictive structural determination approach for small organic molecules.

RESUMEN

Propiedades conformacionales de RMN de ¹H y ¹³C calculadas con Spartan 18 y datos experimentales de 400/125,77 MHz de β-orcinolcarboxilato de metilo y atranorina, un estudio comparativo

Los parámetros estructurales del metil β-orcinolcarboxilato y la atranorina se determinaron mediante B3LYP con el conjunto base 6-31G*. Se presentan los resultados de la estructura molecular optimizada y se comparan con los datos de rayos X disponibles de las moléculas. Se expone y discute un análisis comparativo de los espectros de RMN ¹H y ¹³C experimentales y los valores calculados. Los cálculos cuánticos teóricos dieron como resultado un buen enfoque de determinación estructural predictiva para moléculas orgánicas pequeñas.

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INTRODUCTION

Methyl β -orcinolcarboxylate (**1**) and atranorin (**2**) (Fig. 1) are important natural products isolated from Bolivian Lichens. Atranorin has been characterized in *Stereocaulon strictum*¹ *Stereocaulon ramulosum*², *Stereocaulon tomentosum*³, *Physcia solediosa*⁴; methyl β -orcinolcarboxylate is present in *Stereocaulon ramulosum*². These compounds are reknown for manifesting a wide variety of biological activities, including antioxidant, antibacterial, anticancer, and antiangiogenic^{5 6 7 8 9 10 11 12 13 14} among others. The growing interest in these compounds and their potential use in medical applications is evidenced by the increasing number of publications. Hence, correct structural determination and knowledge about the three-dimensional (3D) atomic structure are essential. NMR experimental data are employed in comparison with electronic and conformational properties of methyl β -orcinolcarboxylate and atranorin resulting from theoretical calculations. In the present comparative study of methyl β -orcinolcarboxylate and atranorin, precise quantum mechanical calculations were carried out, both in vacuo, with the aim of analyzing the conformational equilibria and finding the most stable corresponding equilibrium of the structures. This work is framed in a project that started with the computer-assisted study of ¹H and ¹³C NMR spectra of 4-hydroxy-3-(3'-methyl-2'-butenyl)acetophenone isolated from *Senecio graveolens* and its microwave-assisted synthetic derivate, 4'-hydroxy-3'-(3-methyl-2-butenyl)chalcone¹⁵.

EXPERIMENTAL

NMR system and operating conditions

Spectra were run at 400 MHz and 125.77 MHz for ¹H and ¹³C NMR, respectively, using CDCl₃ as solvent. NMR assignments were done based on ¹H, ¹³C 1D NMR and ¹H-¹³C HSQC and HMBC 2D NMR experiments.

Compound (**1**), White crystals; m.p. 140 °C; ¹H NMR (CDCl₃), (400 MHz); δ 3.9 (3H, *s*, H-10), 2.4 (3H, *s*, H-8), 2.1 (3H, *s*, H-9), 6.2 (1H, *s*, H-5). ¹³C NMR (CDCl₃), (125.77 MHz); δ 172.6, 162.6, 159.5, 139.8, 110.6, 108.9, 104.9, 51.5, 23.8, 7.5.

Compound (**2**), White crystals; m.p. 140 °C; ¹H NMR (CDCl₃), (400 MHz); δ 6.44 (1H, *s*, H-5), 6.56 (1H, *s*, H-5'), 2.71 (3H, *s*, H-8), 10.39 (1H, *s*, H-9), 2.11 (3H, *s*, H-8'), 2.59 (3H, *s*, H-9'), 4.05 (3H, *s*, CH₃O-7'), 12.53 (1H, *s*, OH), 12.59 (1H, *s*, OH). ¹³C NMR (CDCl₃), (125.77 MHz); δ 103.8, 169.3, 108.8, 167.7, 113.1, 152.6, 169.9, 25.7, 194.0, 117.0, 163.1, 110.5, 152.2, 116.2, 140.1, 172.4, 24.2, 9.6, 52.5.

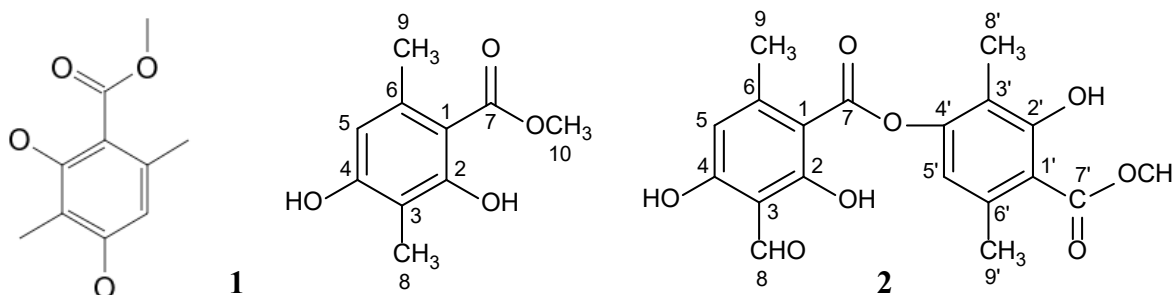


Fig. 1. Methyl β -orcinolcarboxylate (**1**); atranorin (**2**)

Computational details

The geometry of structures was optimized with the DFT methodology using ¹³C NMR chemical shifts of the compounds. In (**1**) each geometrically optimized conformer was calculated using the parameter B3LYP/631G* basis sets. The calculations for (**2**) were carried out in the same way as for (**1**). Molecular mechanics calculations were performed using the Spartan 18 modeling software. Solvent effects were not taken into account in any calculation. From X-ray crystallography data^{16 17} the existence of compounds (**1**) and (**2**) in their solid state was known. In the present work we investigated all possible conformers of methyl β -orcinolcarboxylate and atranorin in order to establish a reliable starting point to further theoretically explore the complexing ability of these compounds.

RESULTS AND DISCUSSION

From X-ray crystallography data^{16 17} a randomized conformational search of the methyl β -orcinolcarboxylate and atranorin was performed. The 8 more significant conformations of methyl β -orcinolcarboxylate and its ¹³C NMR chemical shifts were saved (Fig. 2). Figures 3 and 4 show that the two most significant conformers of methyl β -orcinolcarboxylate, have the intramolecular hydrogen bond between the 2-OH and 7-C=O. They also show the two conformer differences in the 4-OH positions. Conformer I (Fig. 2) exhibits the dihedral angle of 179.24 and in conformer II the angle is 0.26 (Fig. 3). Table 1 shows the calculated (δ_{calc}) and experimental (δ_{exp}) ¹³C-NMR, ¹H-NMR chemical shifts. Data comparison showed agreement between experimental and calculated NMR chemical shifts to an acceptable degree for conformer II. The comparison with image derived from X-ray analysis¹⁶ (Fig.5) ratifies the spectral and calculated data (C₁₀H₁₂O₄, *M_r* = 196.2, Monoclinic, *P*2₁/*c*, *a* = 7.043 (2) Å, *b* = 18.142 (7) Å, *c* = 7.238 (2) Å, β = 96.44 (2)°, *V* = 918.9 (4) Å³, *Z* = 4)¹⁶. The results presented in table 1 and 2 demonstrate an excellent predictive ability of this method for ¹H and ¹³C NMR.

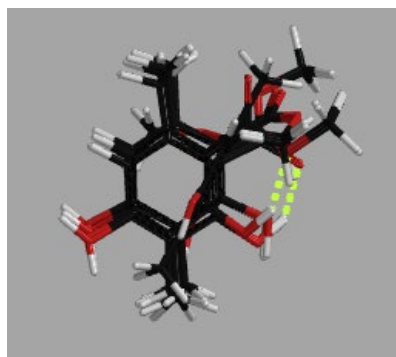


Fig. 2. Eighth Conformers

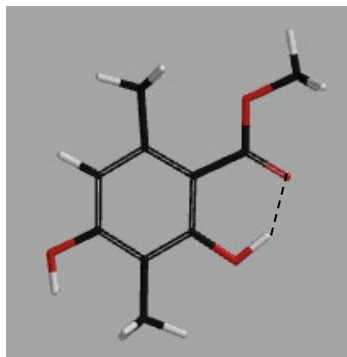


Fig. 3. Conformer I

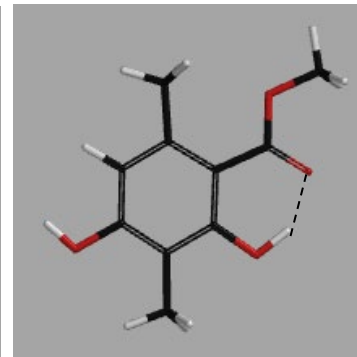


Fig. 4. Conformer II

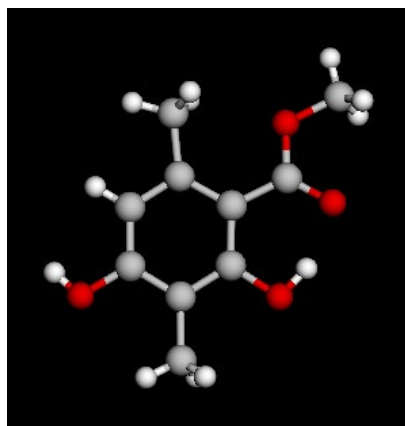


Fig. 5. X-ray crystallography¹⁵

Table 1. Experimental and calculated ¹³C-NMR chemical shifts, CDCl₃, δ from TMS, conformers I and II, compound **1**

Atom		Exp. δ (ppm) ¹³ C	Conformer I	Conformer II
			Calc. δ (ppm) ¹³ C	Calc. δ (ppm) ¹³ C
1	C	104,9	104,2	104,5
2	C	162,6	163,4	165,2
3	C	108,9	105,8	110,7
4	C	159,5	158,3	156,0
5	CH	110,6	110,2	109,2
6	C	139,8	142,0	140,2
7	C=O	172,6	173,6	173,3
8	CH ₃	23,8	25,5	25,3
9	CH ₃	7,5	6,6	10,0
OCH3	CH ₃	51,1	51,9	51,9

Atranorin (**2**) being a more complex molecule than compound **1**, 128 conformers (at least) can be determined out of calculations using our current-use method. Thus, the fully optimized structure of atranorin was obtained at the *B3LYP/6-31G** level. Two conformers were picked out and saved, ^{13}C -NMR and ^1H -NMR chemical shifts were scaled.

Table 2. The experimental and calculated ^1H -NMR chemical shifts, CDCl_3, δ from TMS, conformers I and II, compound **1**

Atom		Exp. δ (ppm) ^1H	Conformer I Calc. δ (ppm) ^1H	Conformer II Calc. δ (ppm) ^1H
1	C			
2	C			
3	C			
4	C			
5	CH	6,2	6,0	5,4
6	C			
7	C=O			
8	CH ₃	2,4	2,3	2,3
9	CH ₃	2,1	1,9	2,0
OCH ₃	CH ₃	3,9	3,8	3,8

Tables 3 and 4 show the calculated (δ_{calc}) and experimental (δ_{exp}) ^{13}C -NMR and ^1H -NMR chemical shifts of compound **2**, respectively. The data comparison demonstrated a great agreement between experimental and calculated NMR chemical shifts for conformers A (Fig. 6) and B (Fig. 7). On the other hand, the comparison of X-ray crystallography structure of Fig. 8,¹⁷ ($\text{C}_{19}\text{H}_{18}\text{O}_8$, $M_r = 374.33$ Monoclinic, $P2_1/n$, $a = 10.929$ (3) Å, $b = 10.976$ (3) Å, $c = 14.843$ (3) Å, $\beta = 109.745$ (12)°, $V = 1675.7$ (7) Å³, $Z=4$)¹⁷ with both conformers shows a good matching with conformer B. These results (Tables 3 and 4) demonstrate an excellent predictive ability of the method for ^1H and ^{13}C NMR data.

The two aromatic rings form a dihedral angle of 60.38°, differing from the previously reported *Pbca* polymorph, *viz.* 84°.⁵ In this work we calculated at the *B3LYP/6-31G** two conformers A and B with the two aromatic rings forming a dihedral angle of 86.95°, different from the reported by X-ray crystallography. Nevertheless there is good agreement with the experimental values of NMR ^1H and ^{13}C .

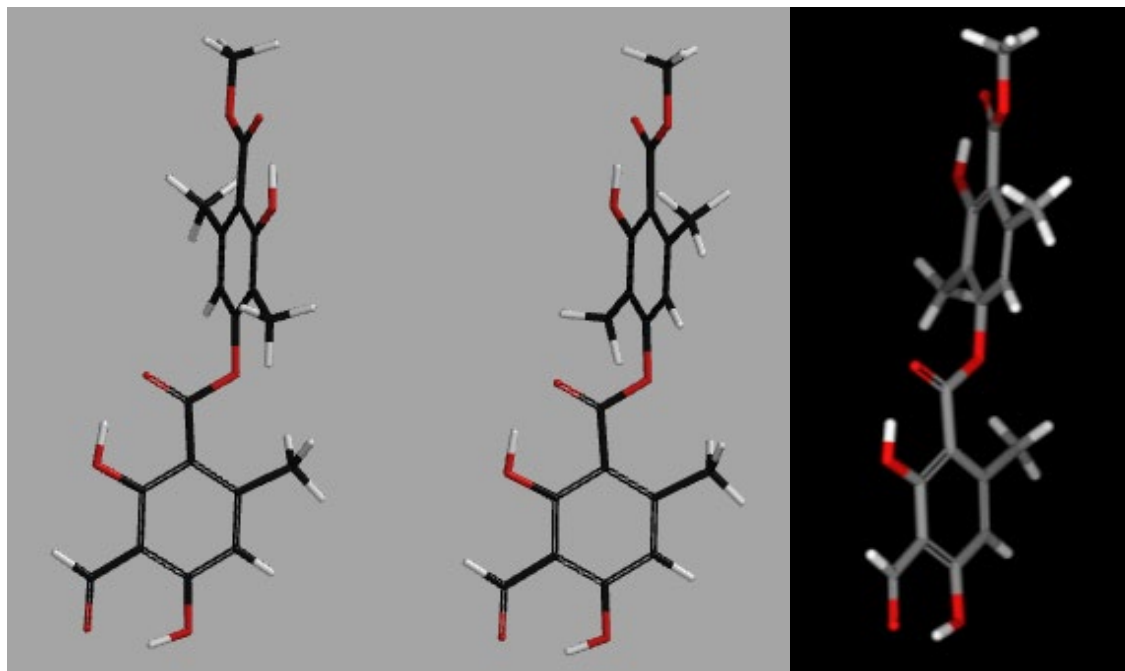


Fig. 6. Conformer A

Fig. 7. Conformer B

Fig. 8. X-ray image¹⁷

Data statistical approach

Plotting the experimental ^{13}C NMR (δ_{exp}) in CHCl_3 vs. the δ_{calc} for all species, a linear regression is obtained. This relationship is used to predict the chemical shifts (δ_{calc}).

Table 3. The experimental and calculated ^{13}C -NMR chemical shifts, CDCl_3 , δ from TMS, compound A and B

Atom		Exp. δ (ppm) ^{13}C	Conformer A Calc. δ (ppm) ^{13}C	Conformer B 5 Calc. δ (ppm) ^{13}C
1	C	103,1	102,1	102,3
2	C	169,3	169,8	169,7
3	C	108,8	109,1	109,1
4	C	167,7	167,5	167,5
5	CH	113,1	113,0	113,1
6	C	152,6	152,0	151,9
7	C=O	169,9	171,5	172,0
9	CH_3	25,7	26,2	25,2
8	CHO	194,0	192,4	192,4
1'	C	110,5	109,0	108,9
2'	C	163,1	164,6	164,9
3'	C	117,0	120,1	119,5
4'	C	152,2	153,5	153,3
5'	CH	116,2	116,5	117,3
6'	C	140,1	140,6	140,4
7'	C=O	172,4	174,0	174,0
8'	CH_3	24,2	25,2	25,2
9'	CH_3	9,6	11,6	11,4
OCH_3	OCH_3	52,5	52,4	52,4

Table 4. The experimental and calculated ^1H -NMR chemical shifts, CDCl_3 , δ from TMS, conformers A and B

Atom		Exp. δ (ppm) ^1H	Conformer A Calc. δ (ppm) ^1H	Conformer B Calc. δ (ppm) ^1H
1	C			
2	C			
3	C			
4	C			
5	CH	6,44	5,9	5,9
6	C			
7	C=O			
9	CH_3	2,71	2,4	2,4
8	CHO	10,39	10,3	10,3
1'	C			
2'	C			
3'	C			
4'	C			
5'	CH	6,56	6,2	6,2
6'	C			
7'	C=O			
8'	CH_3	2,11	2,4	2,4
9'	CH_3	2,59	2,0	2,0
OCH_3	OCH_3	4,05	3,9	3,9

The correlation between experimental chemical shifts and calculated isotropic screening constants for ^{13}C , shows a homogeneous behavior for ^{13}C independently on the calculated species with a correlation coefficient R^2 0.9974 for methyl β -orcinolcarboxylate (Fig. 9) and 0.9966 for atranorin (Fig. 10).

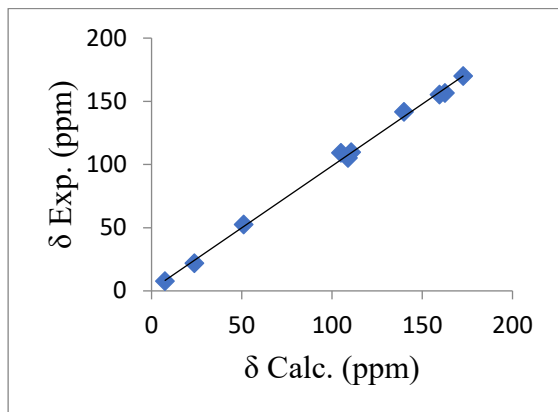


Fig. 9. Experimental chemical shifts of methyl β -ornicolcarboxylate vs. isotropic magnetic from /B3LYP/6-31G* calculations for ^{13}C

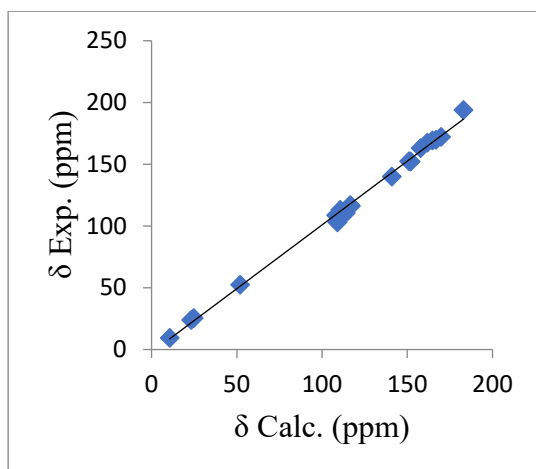


Fig. 10. Experimental chemical shifts of atranorin vs. isotropic magnetic from /B3LYP/6-31G* calculations for ^{13}C

CONCLUSION

The experimental geometric parameters of the compound are comparable to the calculated values. The molecular structure was established by the evidence of short intra-molecular contact interactions. The ^1H and ^{13}C NMR chemical shifts calculations showed an excellent agreement with the experimental data. Overall, the agreement between theoretical and experimental values is agreeable for every structural and spectroscopic calculation. From X-ray bibliography we established the more stable conformers of **1** and **2**, II and B, respectively.

REFERENCES

- ¹ G. Jordán G., 1991, Estudio fitoquímico del líquen *Stereocaulon strictum* (BSc thesis), Universidad Mayor de San Andrés, La Paz, Bolivia.
- ² J. Vila, P. Mollinedo, Y. Flores, and O. Sterner, *Rev. Bol. Quim.*, 2008, **25**, 1.
- ³ J. Vila, P. Canaviri, and O. Sterner, *Rev. Bol. Quim.*, 2004, **21**, 76.
- ⁴ A. Maldonado, R. Meneses, J. A. Bravo and J. L. Vila, *Rev. Bol. Quim.*, 2016, **33**, 174.
- ⁵ B. Ranković, M. Mišić, and S. Sukdolak, The antimicrobial activity of substances derived from the lichens *Physcia aipolia*, *Umbilicaria polyphylla*, *Parmelia caperata* and *Hypogymnia physodes*, *World J. Microbiol. Biotechnol.*, 2008, **24**, 123.
- ⁶ Y. J. Jeon, S. Kim, J. H. Kim, U. J. Youn, and S. S. Suh, The comprehensive roles of Atranorin, a secondary metabolite from the antarctic lichen *Stereocaulon caespitosum*, in HCC tumorigenesis, *Molecules* 2019, **24**, 1414.



- ⁷ M. Bačkorová, M. Bačkor, Mikeš, J. R. Jendželovský, and P. Fedoročko, Variable responses of different human cancer cells to the lichen compounds parietin, atranorin, usnic acid and gyrophoric acid, *Toxicol. Vitr.*, 2011, **25**, 3.
- ⁸ M. Bačkorová, R. Jendželovský, M. Kello, M. Bačkor, J. Mikeš, and P. Fedoročko, Lichen secondary metabolites are responsible for induction of apoptosis in HT-29 and A2780 human cancer cell lines, *Toxicol. Vitr.* 2012, **26**, 462.
- ⁹ V. M. Thadhani, M. A. Mesaik, M. Asif, V. Karunaratne, I. M. Choudhary, Immunomodulatory activities of some common lichen metabolites. *Int. J. Pharm. Pharm. Sci.*, 2015, **7**, 144.
- ¹⁰ T. H. Vu, A. C. Le Lamer, C. Lalli, J. Boustie, M. Samson, F. Lohézic-Le Dévéhat, and J. Le Seyec, Depsides: Lichen metabolites active against hepatitis C virus, *PLoS ONE* 2015, **10**, e0120405.
- ¹¹ S. K. C. Kumar and K. Müller, Lichen metabolites. 1. Inhibitory action against leukotriene B4 biosynthesis by a non-redox mechanism, *J. Nat. Prod.*, 1999, **62**, 817.
- ¹² N. Valencia-Islas, A. Zambrano, and J. Rojas, Ozone reactivity and free radical scavenging behavior of phenolic secondary metabolites in lichens exposed to chronic oxidant air pollution from Mexico City, *J. Chem. Ecol.*, 2007, **33**, 1619.
- ¹³ M. G. D. Melo, A. A. S. Araujo, C. P. L. Rocha, E. M. S. A. Almeida, R. de Souza Siqueira, L. R. Bonjardim, L. J. Quintans-Júnior, Purification, physicochemical properties, thermal analysis and antinociceptive effect of atranorin extracted from *Cladonia kalbi*, *Biol. Pharm. Bull.*, 2008, **31**, 1977.
- ¹⁴ S. Bo, Bactericidal activity of some lichen secondary compounds of *Cladonia ochrochlora*, *Parmotrema nilgherrensis* & *Parmotrema sancti-angelii*, *Int. J. Drug Dev. Res.*, 2011, **3**, 222.
- ¹⁵ J. A. Bravo and J. L. Vila, *Rev. Bol. Quim.*, 2019, **36**, 104.
- ¹⁶ G. J. Gainsford, Antifungal Compounds Isolated from New Zealand Flax: 7-Hydroxy-5-methoxy-6-methylphthalide and 4-Methoxycarbonyl-3-orcinol, *Acta Cryst.*, 1995, **C51**, 709.
- ¹⁷ Y. H. Liu, S. L., Robbs, F. R. Fronczek, S. F. Watkins, and N. H. Fischer, A new polymorph of atranorin, a lichen paradespide, *Acta Cryst.*, 2002, **E58**, o877-o878.